

Prevalence and Clinical Implications of a β -Amyloid-Negative, Tau-Positive Cerebrospinal Fluid Biomarker Profile in Alzheimer Disease

Pontus Erickson, BSc; Joel Simrén, MD; Wagner S. Brum; Gilda E. Ennis, PhD; Gwendlyn Kollmorgen, PhD; Ivonne Suridjan, PhD; Rebecca Langhough, PhD; Erin M. Jonaitis, PhD; Carol A. Van Hulle, PhD; Tobey J. Betthausen, PhD; Cynthia M. Carlsson, MD, MSc; Sanjay Asthana, MD; Nicholas J. Ashton, PhD; Sterling C. Johnson, PhD; Leslie M. Shaw, MD, PhD; Kaj Blennow, MD, PhD; Ulf Andreasson, PhD; Barbara B. Bendlin, PhD; Henrik Zetterberg, MD, PhD; for the ADNI Cohort

 Supplemental content

IMPORTANCE Knowledge is lacking on the prevalence and prognosis of individuals with a β -amyloid-negative, tau-positive (A-T+) cerebrospinal fluid (CSF) biomarker profile.

OBJECTIVE To estimate the prevalence of a CSF A-T+ biomarker profile and investigate its clinical implications.

DESIGN, SETTING, AND PARTICIPANTS This was a retrospective cohort study of the cross-sectional multicenter University of Gothenburg (UGOT) cohort (November 2019-January 2021), the longitudinal multicenter Alzheimer Disease Neuroimaging Initiative (ADNI) cohort (individuals with mild cognitive impairment [MCI] and no cognitive impairment; September 2005-May 2022), and 2 Wisconsin cohorts, Wisconsin Alzheimer Disease Research Center and Wisconsin Registry for Alzheimer Prevention (WISC; individuals without cognitive impairment; February 2007-November 2020). This was a multicenter study, with data collected from referral centers in clinical routine (UGOT) and research settings (ADNI and WISC). Eligible individuals had 1 lumbar puncture (all cohorts), 2 or more cognitive assessments (ADNI and WISC), and imaging (ADNI only) performed on 2 separate occasions. Data were analyzed on August 2022 to April 2023.

EXPOSURES Baseline CSF A β 42/40 and phosphorylated tau (p-tau)181; cognitive tests (ADNI: modified preclinical Alzheimer cognitive composite [mPACC]; WISC: modified 3-test PACC [PACC-3]). Exposures in the ADNI cohort included [18 F]-florbetapir amyloid positron emission tomography (PET), magnetic resonance imaging (MRI), [18 F]-fluorodeoxyglucose PET (FDG-PET), and cross-sectional tau-PET (ADNI: [18 F]-flortaucipir, WISC: [18 F]-MK6240).

MAIN OUTCOMES AND MEASURES Primary outcomes were the prevalence of CSF AT biomarker profiles and continuous longitudinal global cognitive outcome and imaging biomarker trajectories in A-T+ vs A-T- groups. Secondary outcomes included cross-sectional tau-PET.

RESULTS A total of 7679 individuals (mean [SD] age, 71.0 [8.4] years; 4101 male [53%]) were included in the UGOT cohort, 970 individuals (mean [SD] age, 73 [7.0] years; 526 male [54%]) were included in the ADNI cohort, and 519 individuals (mean [SD] age, 60 [7.3] years; 346 female [67%]) were included in the WISC cohort. The prevalence of an A-T+ profile in the UGOT cohort was 4.1% (95% CI, 3.7%-4.6%), being less common than the other patterns. Longitudinally, no significant differences in rates of worsening were observed between A-T+ and A-T- profiles for cognition or imaging biomarkers. Cross-sectionally, A-T+ had similar tau-PET uptake to individuals with an A-T- biomarker profile.

CONCLUSION AND RELEVANCE Results suggest that the CSF A-T+ biomarker profile was found in approximately 5% of lumbar punctures and was not associated with a higher rate of cognitive decline or biomarker signs of disease progression compared with biomarker-negative individuals.

Author Affiliations: Author affiliations are listed at the end of this article.

Group Information: The members of the ADNI Cohort appear in Supplement 2.

Corresponding Authors: Henrik Zetterberg, MD, PhD (henrik.zetterberg@clinchem.gu.se), and Pontus Erickson, BSc (pontus.eriksson@gu.se), Institute of Neuroscience and Physiology, Department of Psychiatry and Neurochemistry, Sahlgrenska Academy, University of Gothenburg, Medicinaregatan 11, 403 30 Gothenburg, Sweden.

JAMA Neurol. 2023;80(9):969-979. doi:10.1001/jamaneurol.2023.2338
Published online July 31, 2023.

Cerebrospinal fluid (CSF) analysis is a well-established method to support a clinical diagnosis of Alzheimer disease (AD), with validated and clinically approved biomarkers for β -amyloid ($A\beta$) pathology, phosphorylated tau (p-tau), and neurodegeneration. In recent research criteria, low $A\beta_{42}$ or $A\beta_{42}/40$ ratio and increased concentration of p-tau have been proposed to support the diagnosis of AD,¹ with the AT(N) system classifying CSF $A\beta$ and p-tau in the “A” and “T” categories, respectively.² A proposed model of AD pathogenesis suggests that biomarkers reflecting $A\beta$ pathology become abnormal before measures of tau pathology, which is now supported by a vast body of evidence.³

Normal CSF $A\beta$ in combination with increased p-tau is a finding sometimes seen in clinical settings.² Because p-tau increases are suggested to be AD specific,^{4,5} and increases would not be expected in $A\beta$ -negative individuals, this finding is challenging to interpret. Guidelines vary in the terminology assigned to this finding, mostly encompassing the concept of suspected non-Alzheimer pathology.⁶ For example, studies evaluating the prevalence of CSF biomarker abnormalities suggest that such an A-T+ CSF profile can be found in up to 20% of cognitively unimpaired (CU) older adults.⁷⁻¹⁰ In addition, available studies show mixed results in the progression of disease-related changes in individuals with this biomarker profile.^{7,10,11} Previously, various propositions have been made to explain its underlying cause, including non-AD tauopathies (ie, primary age-related tauopathy) and altered CSF dynamics, among others.^{6,12} Importantly, these explanations likely vary depending on the biomarker modality being used. Unlike $A\beta$ biomarkers, which seem to provide similar information in terms of “A” status when determined by positron emission tomography (PET) or CSF, tau biomarkers present pronounced differences across imaging and fluid modalities. Although fibrillar tau aggregates targeted by tau-PET radiotracers are likely a more direct proxy of tangle pathology,^{13,14} soluble p-tau measured by immunoassays may reflect a neuronal response to $A\beta$ pathology,¹⁵⁻¹⁸ although p-tau is also present in dystrophic neurites, suggesting that it also reflects tau pathology.^{19,20} Although evidence is accumulating that individuals with an A-T+ biomarker profile (and to a greater extent, those with both CSF signs of $A\beta$ and soluble tau pathology [A+T+]) are at risk for cognitive decline,^{7,10} less is known about the clinical implications of the CSF A-T+ pattern. Because CSF biomarkers are frequently used in clinical practice, it is important to further understand the implications of patterns not expected within the currently proposed model of AD biomarker progression.

Given the lack of studies directly evaluating the clinical relevance and prevalence of an isolated increase in CSF p-tau, we aimed to assess the prevalence, as well as cognitive and biomarker progression rates, associated with the CSF A-T+ profile, using 4 large data sets.

Methods

Design

Four cohorts were used in the present study: University of Gothenburg (UGOT), the Alzheimer Disease Neuroimaging Initiative (ADNI), Wisconsin Registry for Alzheimer Preven-

Key Points

Question What is the memory clinic prevalence and prognosis of individuals who have a β -amyloid-negative, tau-positive (A-T+) cerebrospinal fluid (CSF) profile?

Findings In a clinical cohort of 7679 individuals, prevalence of A-T+ CSF was 4.1%. Longitudinally, cognitively unimpaired individuals or individuals with mild cognitive impairment and an A-T+ CSF profile exhibited similar trajectories to individuals with an A-T- CSF profile in relation to cognitive deterioration, brain atrophy, or cerebral glucose metabolism and β -amyloid burden indexed by positron emission tomography.

Meaning A CSF profile of A-T+ appears to be benign despite being classified as a pathologic change by guidelines; compared with individuals with biomarker-negative CSF, individuals with A-T+ CSF do not have higher rates of cognitive decline or faster disease progression.

tion (WRAP), and Wisconsin Alzheimer Disease Research Center (WI-ADRC). For each cohort, a local institutional review board for human research or ethics committee approved of the study. The UGOT cohort consisted of cross-sectional data and was retrospectively compiled using anonymized data from the clinical routine laboratory database at Sahlgrenska University Hospital, Gothenburg, Sweden, based on CSF analyses performed between November 2019 and January 2021. ADNI is a longitudinal multicenter observational study with data being collected between September 2005 and May 2022. WRAP and the Wisconsin Alzheimer Disease Research Center (WI-ADRC) are single-center, longitudinal, observational, cohort studies that collected data between February 2007 and November 2020. The present study followed the Strengthening the Reporting of Observational Studies in Epidemiology (STROBE) reporting guidelines.

Setting

The UGOT cohort consists of individuals who underwent lumbar puncture primarily in memory and neurology clinics. The ADNI is a study of volunteers with MCI or early AD, as well as CU individuals enrolled at memory clinics.¹¹ The WISC sample included longitudinally observed participants enrolled at midlife at the WRAP or the WI-ADRC referral centers.²¹ Due to their similarities in data collection, WRAP and WI-ADRC were combined and analyzed together and will subsequently be denoted as WISC.

Participants

In the UGOT cohort, all participants were older than 50 years and had data on age, sex, and CSF biomarker concentrations, whereas clinical information on diagnosis and cognitive function was not available. In the well-characterized longitudinal observational cohorts (ADNI and WISC), we included only individuals who were CU or had mild cognitive impairment (MCI; only in ADNI) at the first cognitive evaluation. In the UGOT cohort, no data on participant race and ethnicity were available, as they are not routinely collected by health care services. In the ADNI cohort, race categories included were

American Indian or Alaska Native, Asian, Black, Hawaiian, multiracial, and White. In the WISC cohort, individuals self-identifying as White were reported, whereas American Indian or Alaska Native, Asian, Black or African American, or those not identifying as any of the stated races (here indicated as other), were not reported to maintain anonymity for groups with less than 3 individuals. More information on all cohorts used can be found in eMethods 1 in [Supplement 1](#), and detailed inclusion and exclusion criteria have been described for the ADNI²² and WISC cohorts.²¹ Written informed consent was obtained from participants in the ADNI and WISC cohorts, and local institutional review boards for human research approved the study. For the UGOT data, in accordance with the Helsinki declaration, an external ethics committee has approved the procedure of using data from this cohort in research purposes.⁵ There were no identifiable data used in this study, and thus, no informed consent was needed.

CSF Biomarkers

CSF A β 42/40 ratio and p-tau181 (hereafter referred to as p-tau) were quantified in all participants and were used to define AT status in all 3 cohorts. In the UGOT cohort, CSF concentrations of A β 42, A β 40, and p-tau were quantified on the Lumipulse G1200 platform (Fujirebio), a fully automated assay recently approved by the US Food and Drug Administration.²³ In both the ADNI and WISC cohorts, p-tau was quantified using a fully automated Elecsys assay (Roche), also recently approved by the US Food and Drug Administration. A β 42 and A β 40 were measured with mass spectrometry and Elecsys assays in the ADNI and WISC cohorts, respectively. Elecsys A β 42 is an in vitro diagnostic test approved for use in the European Union and US; A β 40 was measured using a research use-only Elecsys assay. CSF status determined at baseline lumbar puncture was tested in the WISC analyses. In all cohorts, previously established cohort-specific cutoffs were used. In ADNI, cutoffs of 0.0138 or less and 24 pg/mL or greater were used for A β 42/40 and p-tau, respectively. In WISC, abnormal biomarker results were determined to be 0.046 or less and 24.8 pg/mL or greater for A β 42/40 and p-tau, respectively. More information on the CSF collection and quantification of and cutoff determination for A β 42, A β 40, and p-tau can be found in eMethods 2 in [Supplement 1](#).

Imaging Biomarkers

In longitudinal imaging analyses in ADNI participants, we included those who had at least a baseline and a follow-up scan. We evaluated longitudinal trajectories of A β [18 F]-florbetapir, [18 F]-fluorodeoxyglucose (FDG) PET, and hippocampal volume magnetic resonance imaging (MRI). A brief description of imaging methods can be found in eMethods 3 in [Supplement 1](#).^{24,25} As a secondary analysis, we evaluated summary tau-PET measures between CSF AT groups, to investigate whether elevated p-tau in the absence of A β could represent an abnormal tau-PET phenotype. In the WISC sample, a subset of participants underwent [18 F]-MK6240 tau-PET and were evaluated with an entorhinal cortex region of interest (ROI), as further detailed in eMethods 3 in [Supplement 1](#).²⁶

In the ADNI cohort, a subsample of the individuals herein included had available [18 F]-flortaucipir tau-PET data up to 6 years after the baseline lumbar puncture, and the previously described ADNI meta-temporal ROI was used as the composite region.²⁷

Cognitive Assessments

The cognitive measures used for longitudinal analyses in the 2 observational cohorts were 2 different versions of the pre-clinical Alzheimer cognitive composite (PACC): the modified PACC (mPACC) in ADNI and a modified 3-test PACC (PACC-3) in WISC.²⁸⁻³⁰ The PACC is a sensitive measure for tracking early cognitive decline, composed typically of 3 key domains: episodic memory, executive function, and global cognition.^{31,32} Specific mPACC and PACC-3 details can be found in eMethods 4 in [Supplement 1](#).

Statistical Analysis

In the UGOT cohort, we estimated the prevalence of the CSF AT profiles across ages. To better visualize the prevalence across different ages, we fitted a locally estimated scatterplot smoothing line (LOESS), which is a flexible nonlinear regression approach. In all cohorts, general prevalence of each CSF AT profile was calculated, and 95% CIs were estimated using 1-sample proportions tests. In the UGOT cohort, we also performed a sensitivity analysis to evaluate if measured concentrations changed over time or if cutoff-related differences influenced the prevalence (eMethods 5 in [Supplement 1](#)).

We used linear mixed-effects models to investigate whether CSF AT group modified the longitudinal rate of change in cognitive function in ADNI (mPACC) and WISC (PACC-3), as well as in A β -PET, FDG-PET, and MRI-derived hippocampal volume (ADNI only). The models were fitted with random slopes and intercepts on the participant level and included CSF AT group (A–T– as reference), age, sex, APOE ϵ 4 carrier status, and years of education, and an interaction term between CSF AT group and time. In the cognition models, a continuous term adjusting for practice effects (ie, number of occasions the outcome tests were taken before last cognitive assessment) was included in both the ADNI and WISC cohorts, with the time being modeled as age. The WISC model also adjusted for cohort membership. In the ADNI cohort, for each outcome, we fitted separate models for CU individuals at baseline and MCI at baseline. To investigate if a more stringent p-tau cutoff influenced the results of the linear mixed-effects models in the ADNI and WISC cohorts, we performed a sensitivity analysis increasing the predetermined p-tau cutoff values by 15%. As a secondary analysis, we cross-sectionally examined differences in tau-PET uptake across CSF AT groups, in the ADNI and WISC cohorts, with linear models, controlling for age, sex, APOE ϵ 4 status, and years of education. Pairwise comparisons using Šidák correction to control for family-wise error rate were performed following a significant omnibus test result. *P* values were 2-sided, and the cutoff for significance was set to *P* < .05. Statistical analyses were performed using R software, version 4.1.2 (R Project for Statistical Computing). Data were analyzed on April 2022 to April 2023.

Results

Baseline CSF Characteristics

A total of 7679 individuals (mean [SD] age, 71.0 [8.4] years; 4101 male [53%]; 3578 female [47%]) were included in the UGOT cohort, 970 individuals (mean [SD] age, 73 [7.0] years; 526 male [54%]; 444 female [46%]) were included in the ADNI cohort, and 519 individuals (mean [SD] age, 60 [7.3] years; 346 female [67%]; 173 male [33%]) were included in the WISC cohort. The ADNI cohort included 364 CU individuals and 606 individuals with MCI. The WISC cohort included 513 CU individuals and 6 individuals with MCI. In the ADNI cohort, study participants identified with the following race and ethnicity categories: 1 American Indian or Alaska Native (0.3%), 9 Asian (2.3%), 25 Black (6.5%), 1 Hawaiian (0.3%), 7 multiracial (1.8%), and 341 White (88%). In the WISC cohort, 493 individuals (95%) self-identifying as White were reported; individuals identifying as American Indian or Alaska Native, Asian, Black or African American, or other were not reported in order to maintain anonymity for groups with 3 or fewer individuals. Key information on demographic and other variables is displayed in the Table.

Prevalence of CSF AT Profiles in a Real-World Clinical Routine Setting

In the UGOT cohort, 316 of 7679 individuals (4.1%; 95% CI, 3.7%-4.6%) displayed an A-T+ profile, whereas the prevalence of A-T- was 3241 of 7679 (42%; 95% CI, 41%-43%), the prevalence of A+T- was 1324 of 7679 (17%; 95% CI, 16%-18%), and the prevalence of A+T+ was 2798 of 7679 (36%; 95% CI, 35%-38%). Similar estimates were found in the ADNI and WISC cohorts (Table). Alternatively, if determining "A" by CSF A β 42 alone, as done in several studies discussed subsequently, the prevalence of an A-T+ profile would be 19% (95% CI, 18%-20%) and 37% (95% CI, 36%-38%), 22% (95% CI, 21%-23%), and 21% (95% CI, 20%-22%) for A-T-, A+T-, and A+T+ profiles, respectively.

In visual interpretation of age-stratified LOESS curves, we found that although the prevalence of other CSF profiles changed dynamically toward abnormality with aging, whereas the prevalence of the A-T+ profile was stable with older age, with a minor increase in the age prevalence after 75 years (Figure 1A). In contrast, when performing the same analysis with A β 42 alone as "A," a different trend was observed. The age-stratified rates of an A-T+ profile displayed a similar trend to that of A+T- and A+T+ profiles, with all presenting increased prevalence with increased age, reaching similar figures of approximately 25% each in the oldest individuals (eFigure 1 in Supplement 1). In a grayscale analysis to evaluate potential analytical effects with more stringent cutoff values and variation in analytical performance over time, no such significant changes were observed, and very minor changes were found in the prevalence of the A-T+ group (eTable 1 and eFigure 2 in Supplement 1). Interestingly, in both the UGOT and ADNI cohorts, concentrations of CSF A β 42 and A β 40 alone were substantially higher in the A-T+ group compared with A-T- (Figure 1B-E).

Cognitive Trajectories

In Figure 2, we display predicted longitudinal cognitive trajectories according to baseline CSF AT profiles. In the ADNI cohort, A-T+ individuals did not present with significantly different rates of decline in mPACC as compared with A-T- individuals, regardless of baseline cognitive status (CU: β = -0.09; 95% CI, -0.3 to 0.1; P = .27; MCI: β = -0.01; 95% CI, -0.4 to 0.4; P = .99). In the WISC cohort, which included predominantly CU individuals, the A-T+ profile did not show significantly different rates of decline in PACC-3 scores compared with A-T- individuals (β = -0.02; 95% CI, -0.05 to 0.02; P = .28). In both the ADNI (CU and MCI) and WISC (CU at baseline) cohorts, individuals with A+T- and A+T+ profiles had faster cognitive decline when compared with those with an A-T- profile (eTables 2 and 3 in Supplement 1). When using more stringent p-tau cutoff values, similar results were obtained. Full models from both main and sensitivity analyses are available in eTables 2, 3, 4, and 5 in Supplement 1.

Change in Imaging Markers

Next, longitudinal imaging biomarkers were used to assess disease progression over time (Figure 3). In the ADNI cohort, regardless of their baseline cognitive status, individuals with an A-T+ profile did not differ significantly from those with A-T- in longitudinal rates of change in A β -PET (CU: β = 0.002; 95% CI, -0.01 to 0.01; P = .44; MCI: β = 0.002; 95% CI, -0.01 to 0.01; P = .44), FDG-PET (CU: β = -0.002; 95% CI, -0.01 to 0.001; P = .28, MCI: β = -0.002; 95% CI, -0.005 to 0.002; P = .39), or hippocampal volume (CU: β = -19 mm³ per year; 95% CI, -40 to 1.0; P = .06; MCI: β = 3.9 mm³ per year; 95% CI, -29 to 37; P = .82), although numerically lower in CU individuals. In contrast and in accordance with previous studies, individuals with MCI and A+T+ or A+T- profiles, when compared with those with an A-T- profile, displayed significant changes in markers of hippocampal volume and glucose metabolism (FDG PET and MRI, respectively) (eTables 6 and 7 in Supplement 1). In CU with an A+T- profile, no significant changes were seen in change in FDG PET, whereas there was a slight decrease in hippocampal volume as compared with individuals with an A-T- profile (eTables 6 and 7 in Supplement 1). However, significant time-related increases in A β -PET were seen for individuals with both A+T- and A+T+ profiles independent of baseline cognitive status. Changing the cutoff values for p-tau yielded very similar results. Full models from both main and sensitivity analyses are available in eTables 6, 7, 8, 9, 10, and 11 in Supplement 1.

Fibrillar Tau Deposition

In a subset of participants who had tau-PET data (ADNI: n = 192; WISC: n = 227), we evaluated whether isolated p-tau positivity was associated with the deposition of fibrillar tau (Figure 4). In the ADNI cohort, individuals with available tau-PET in the A-T+ CSF group did not present statistically significant differences in [¹⁸F]-flortaucipir meta-temporal ROI standardized uptake value ratio (SUVR) values

Table. Baseline Demographics and Clinical Characteristics

Cohort	Biomarker profile				
	All	A–T–	A+T–	A+T+	A–T+
UGOT					
Participants, No. (%); 95% CI, % ^a	7679	3241 (42); 41 to 43	1324 (17); 16 to 18	2798 (36); 35 to 38	316 (4.1); 3.7 to 4.6
Age, mean (SD), y	71 (8.4)	69 (9)	73 (7.6)	73 (7.5)	73 (8.7)
Sex, No. (%)					
Female	3578 (47)	1371 (42)	597 (45)	1487 (53)	123 (39)
Male	4101 (53)	1870 (58)	727 (55)	1311 (47)	193 (61)
CSF A β 42/40, mean (SD)	0.7 (0.62)	0.93 (0.12)	0.57 (0.093)	0.44 (0.094)	1.1 (2.8)
CSF A β 42, mean (SD), ng/L	660 (353)	803 (340)	438 (175)	515 (193)	1410 (457)
CSF A β 40, mean (SD), ng/L	9820 (3920)	8620 (3380)	7680 (2720)	11 700 (3650)	15 000 (4230)
CSF p-tau181, mean (SD), ng/L	56 (44)	28 (9.1)	36 (8.6)	97 (48)	66 (19)
ADNI					
Participants, No. (%); 95% CI, % ^a	970	386 (40); 37 to 43	165 (17); 15 to 20	338 (35); 32 to 38	81 (8.4); 6.7 to 10
Age at baseline, mean (SD), y	73 (7.0)	71 (6.9)	74 (6.6)	74 (7.0)	73 (7.3)
Sex (female), No. (%)					
Female	444 (46)	178 (46)	68 (41)	159 (47)	39 (48)
Male	526 (54)	208 (54)	97 (59)	179 (53)	42 (52)
Race, No. (%) ^b					
American Indian/Alaska Native	1 (0.3)	0	0	0	1 (0.1)
Asian	9 (2.3)	1 (0.6)	2 (0.6)	0	12 (1.2)
Black	25 (6.5)	4 (2.4)	8 (2.4)	1 (1.2)	38 (3.9)
Hawaiian	1 (0.3)	0	1 (0.3)	0	2 (0.2)
Multiracial	7 (1.8)	4 (2.4)	1 (0.3)	0	12 (1.2)
White	341 (88)	156 (95)	326 (96)	80 (99)	903 (93)
MCI, No. (%)	606 (63)	199 (52)	105 (64)	268 (79)	34 (42)
APOE ϵ 4 positive, No. (%) ^c	403 (42)	76 (20)	87 (53)	225 (67)	15 (19)
Education, mean (SD), y	16.2 (2.7)	16.4 (2.7)	15.9 (2.8)	16.0 (2.7)	16.5 (2.7)
CSF A β 42/40, mean (SD)	0.14 (0.057)	0.20 (0.029)	0.11 (0.023)	0.088 (0.021)	0.19 (0.034)
CSF A β 42, mean (SD), ng/L	1180 (620)	1500 (509)	725 (273)	794 (280)	2190 (581)
CSF A β 40, mean (SD), ng/L	8310 (2460)	7640 (2110)	6820 (1840)	9080 (2270)	11300 (2270)
CSF p-tau181, mean (SD), ng/L	256 (13)	16 (3.9)	19 (3.3)	39 (13)	29 (4.8)
mPACC, mean (SD)	–3.5 (4.3)	–1.7 (3.4)	–3.7 (4.2)	–5.9 (4.3)	–1.9 (3.5)
WISC^d					
Participants, No. (%); 95% CI, % ^a	519	412 (79); 76 to 82	49 (9.4); 7.2 to 12	37 (7.1); 5.2 to 9.7	21 (4.0); 2.6 to 6.1
WRAP, No. (%)	233 (45)	178 (34)	27 (5.2)	17 (3.3)	11 (2.1)
WI-ADRC, No. (%)	286 (55)	234 (45)	22 (4.2)	20 (3.8)	10 (1.9)
Age, mean (SD), y	60 (7.3)	59 (6.9)	63 (6.2)	66 (7.8)	65 (8.6)
Age at baseline LP, y	62 (7.5)	61 (7.1)	65 (5.9)	69 (7.2)	68 (8.6)
Sex, No. (%)					
Female	346 (67)	275 (67)	31 (63)	23 (62)	17 (81)
Male	173 (33)	137 (33)	18 (37)	14 (38)	4 (19)
Race, No. (%) ^b					
American Indian/Alaska Native	NR	NR	NR	NR	NR
Asian	NR	NR	NR	NR	NR
Black/African American	NR	NR	NR	NR	NR
White	493 (95)	390 (95)	47 (96)	35 (95)	21 (100)
Other	NR	NR	NR	NR	NR
MCI at baseline LP, No. (%)	6 (1.2)	3 (0.7)	0 (0.0)	1 (2.7)	2 (9.5)
APOE ϵ 4 positive, No. (%) ^c	194 (37)	138 (34)	30 (61)	23 (62)	3 (14)
Education, mean (SD), y	16.2 (2.4)	16.2 (2.5)	15.8 (2.4)	16.6 (2.2)	16.4 (1.9)
CSF A β 42/40, mean (SD)	0.06 (.02)	0.07 (.01)	0.04 (.01)	0.03 (.01)	0.06 (.01)

(continued)

Table. Baseline Demographics and Clinical Characteristics (continued)

Cohort	Biomarker profile				
	All	A-T-	A-T+	A+T+	A-T+
CSF A β 42, mean (SD), ng/L	927 (379)	977 (347)	495 (148)	608 (226)	1513 (195)
CSF A β 40, mean (SD), ng/L	14 421 (4714)	13 786 (4241)	12 947 (3474)	18 128 (3977)	23 794 (4146)
CSF p-tau181, mean (SD), ng/L	18 (6.9)	15 (4.3)	19 (3.9)	32 (7.4)	31 (5.6)
PACC-3, mean (SD)	0.09 (.94)	0.13 (.93)	-0.02 (1.02)	-0.12 (.89)	-0.01 (.93)

Abbreviations: A β 42/40, β -amyloid 1-42 to β -amyloid 1-40; CSF, cerebrospinal fluid; LP, lumbar puncture; MCI, mild cognitive impairment; mPACC, modified preclinical Alzheimer cognitive composite; NR, not reported; p-tau, phosphorylated tau; PACC-3, modified 3-test preclinical Alzheimer cognitive composite.

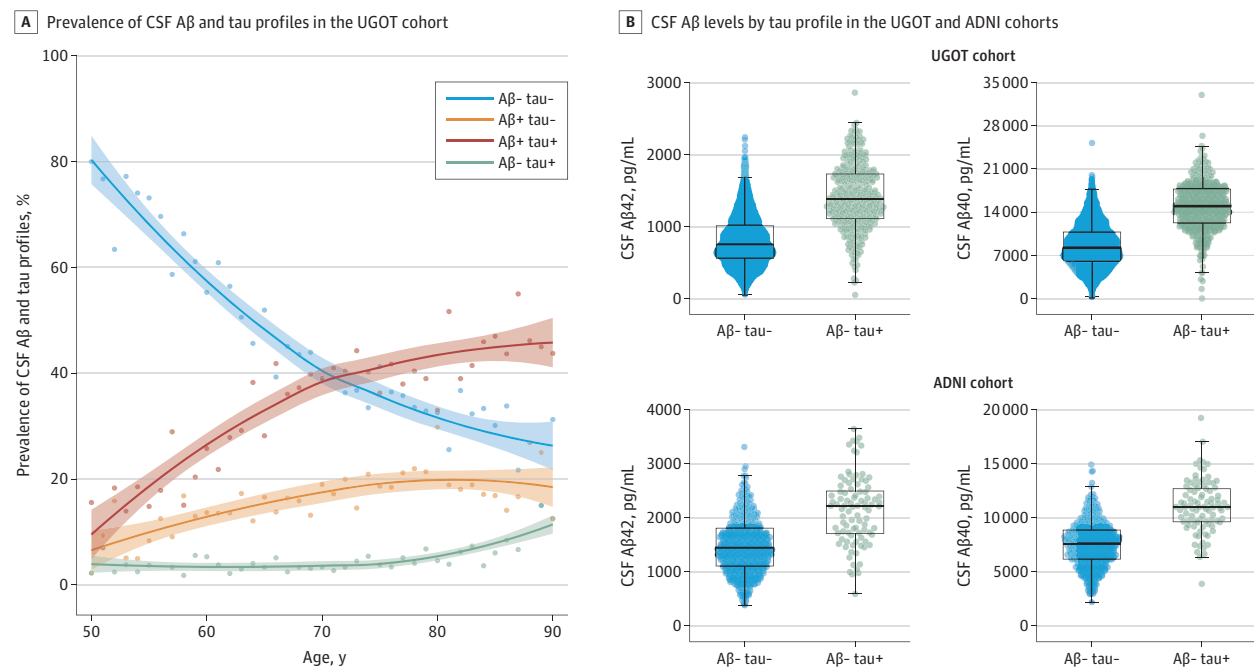
^a 95% CIs were estimated using a 1-sample proportions test.

^b Participants who did not self-identify as White identified as American Indian or

Alaska Native, Asian, Black or African American, or other. Numbers are NR to maintain anonymity for groups with fewer than 3 individuals.

^c Indicates the proportion of individuals carrying at least 1 copy of the *APOE* ϵ 4 allele.

^d In the WISC cohort, demographic and clinical characteristics, except where otherwise noted, are from time point of first cognitive evaluation. CSF data are from the time point of first LP.

Figure 1. Prevalence Estimates in a Clinical Laboratory Routine Setting Across Ages and Cerebrospinal Fluid (CSF) β -Amyloid 42 (A β 42) and A β 40 Concentrations Among Amyloid-Negative Individuals

A, The color dots represent the prevalence in percentage at each age (in years) of each biomarker category based on the cutoffs used in clinical routine. The solid lines represent corresponding locally estimated scatterplot smoothing (LOESS) regression lines, with shaded areas indicating 95% CIs. B, The graphs display group comparisons A β 42 and A β 40 for the A β -negative amyloid-tau (AT) biomarker profiles in both the University of Gothenburg (UGOT) and Alzheimer's Disease Neuroimaging Initiative (ADNI) cohorts. Group comparisons were performed with linear regression models adjusting for relevant available covariates (UGOT: age, sex; ADNI: age, sex, *APOE* ϵ 4 status,

years of education). In the UGOT cohort, mean levels of CSF A β 42 were significantly increased in individuals with an A-T+ profile by a mean of 627 pg/mL (95% CI, 586-667 pg/mL; $P < .001$), and CSF levels of A β 40 were significantly increased in the A-T+ group by a mean of 6504 pg/mL (95% CI, 6104-6904 pg/mL; $P < .001$). In ADNI, mean levels of CSF A β 42 were significantly increased in individuals with an A-T+ profile by a mean of 673 pg/mL (95% CI, 554-793 pg/mL; $P < .001$), and CSF levels of A β 40 were significantly increased in the A-T+ group by a mean of 3543 pg/mL (95% CI, 3054-4054 pg/mL; $P < .001$).

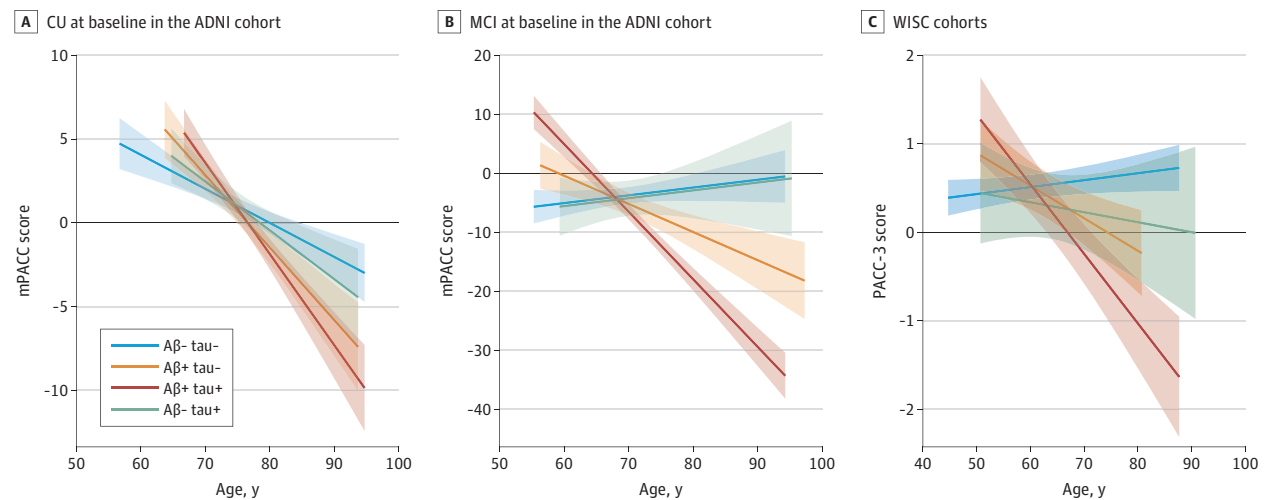
(estimated marginal mean [EM] = 1.2; SE = 0.06) in comparison with the A-T- group ($P = .40$) (EM = 1.2; SE = 0.03). In the WISC cohort, the A-T+ group (EM = 1.1; SE = 0.12) did not show significantly different [18 F]-MK6240 tau-PET SUVR than the A-T- group (EM = 1.1; SE = 0.20; $P = .99$). In both the ADNI and WISC cohorts, the A-T+ group had higher tau PET SUVR (ADNI: EM = 1.54; SE = 0.03; $P < .001$; WISC: EM = 1.5; SE = 0.04; $P = .002$) compared with the A-T+

group. Differences were less clear for the CSF A-T-group (eTables 12 and 13 in Supplement 1).

Discussion

In this cohort study, we found that the prevalence of CSF A-T+ in a clinical neurochemistry laboratory, likely to be represen-

Figure 2. Associations Between Cerebrospinal Fluid (CSF) Amyloid-Tau (AT) Status and Longitudinal Cognitive Decline



Mean predicted trajectories of cognitive decline according to CSF AT status and baseline cognitive status are shown for individuals who were cognitively unimpaired (CU) (A) or had mild cognitive impairment (MCI) (B) at baseline in the Alzheimer's Disease Neuroimaging Initiative (ADNI) and in 2 Wisconsin cohorts, Wisconsin Alzheimer Disease Research Center and Wisconsin Registry for Alzheimer Prevention (WISC) (C). The mean predicted trajectories for the

modified Preclinical Alzheimer's Cognitive Composite (mPACC; ADNI) and modified 3-test PACC (PACC-3; WISC) are displayed with solid lines and 95% CIs. Trajectories were estimated including terms for CSF AT status, covariates (age, years of education, *APOE* $\epsilon 4$ genotype, sex, and practice [and cohort in WISC]), as well as the age \times CSF AT status interaction. A+/- indicates CSF A β 42/40 binary status, and T+/- indicates CSF p-tau181 binary status.

tative of a general memory clinic population, was 4.1%, which is far lower than the prevalence of other CSF profiles. Further, compared with the A-T- biomarker profile, we found that the A-T+ profile was not associated with higher rates of cognitive deterioration, A β accumulation, tau PET pathology, or neurodegeneration.

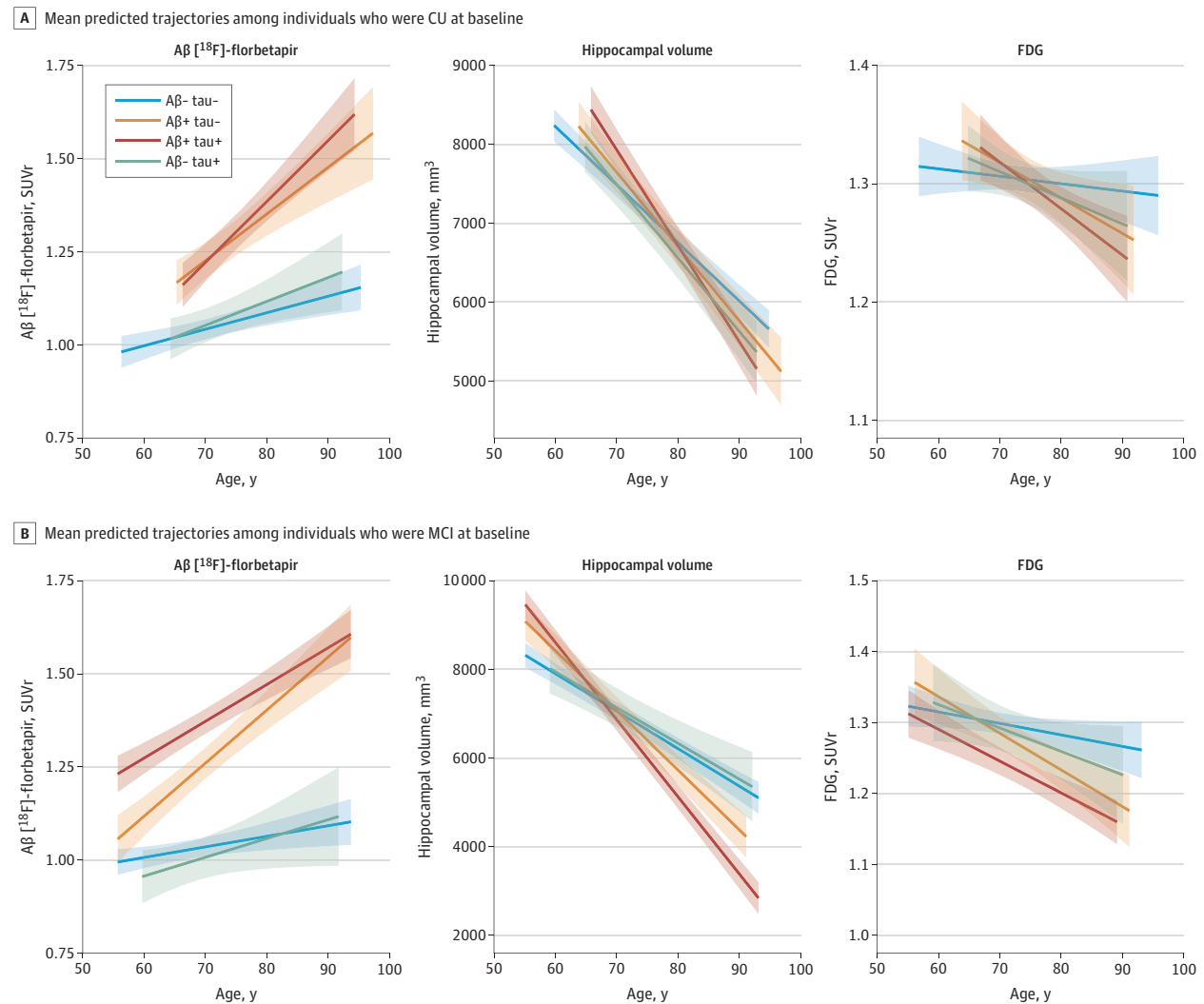
Several studies have attempted to determine the prevalence of AD pathophysiology in varying scenarios. We found that in the UGOT cohort, CSF A β abnormalities ranged from approximately 25% in the youngest individuals to roughly 60% in the oldest individuals. These estimates are lower than those found in a recently published, large, multicenter study evaluating the prevalence of A β positivity as determined by either PET or CSF.³³ This slight discrepancy may be explained by the presence of other disorders causing cognitive symptoms in our cohort and likely reflects more similarly what may be expected in a real-world memory clinic, considering most CSF analyses are ordered by memory clinics or general neurology centers across Sweden. However, fewer studies have performed in-depth evaluations of the clinical meaning of CSF results not consistent with the current view of the chain of AD pathophysiological events. Previous studies evaluating the prevalence of CSF AT abnormalities report mixed results. Although many studies report higher figures (above approximately 15%) for the prevalence of an A-T+ profile, most of these have relied on CSF A β 42 alone to define "A" status.^{7,9,10,34,35} When defining "A" with the widely recommended A β 42/40 ratio,³⁶ this prevalence is substantially reduced to figures like the one reported here for this large, unselected clinical chemistry cohort (approximately 4%).⁸ This suggests that the absence of correction for A β 40 concentrations may lead to artificially deflated frequencies for CSF A β abnormalities, and, consequently, artificially inflated frequen-

cies for a CSF A-T+ profile. This highlights the need for using CSF A β 42/40 in clinical settings when separately assessing "A" and "T" CSF biomarkers.

Furthermore, discussing the etiology underlying a CSF A-T+ profile is highly needed. For instance, individuals presenting with this profile are sometimes referred to as suspected non-Alzheimer's pathology. This construct is most often biomarker defined as A β negativity in the presence of neurodegeneration and a clinical picture not clearly consistent with a recognizable non-AD phenotype.⁶ More specifically, during the early phases of AD CSF biomarker research, p-tau was considered by some studies as a biomarker of neuronal injury.¹⁰ However, it is now known that biomarker abnormalities, especially in "T" and "N" (for neurodegeneration) categories, do not provide the same information within and between modalities.³⁵ Some authors argue that a CSF A-T+ pattern may reflect tau pathophysiology not attributable to A β accumulation, such as primary age-related tauopathy, but this theory has become increasingly untenable due to the accumulation of fluid biomarker evidence that tangle pathology in the absence of concurrent A β pathology is not well reflected by p-tau,^{37,38} although data specifically addressing this question are still scarce. Importantly, we found that the A-T+ group had similar tracer retention in temporal areas as the A-T- group, suggesting that an isolated elevation of CSF p-tau is not due to fibrillar tau pathology in the absence of A β pathology. However, it is important to note that to date, no convincing evidence has shown that tau-PET tracers are capable of capturing PART, which remains a lingering question for the field.

In this challenging context of determining the underpinnings of a CSF A-T+ profile, a potentially better-suited explanation is that the finding of increased p-tau in the absence of

Figure 3. Associations Between Cerebrospinal Fluid (CSF) Amyloid-Tau (AT) Status and Longitudinal Biomarker Signs of β -Amyloid ($A\beta$) Accumulation and Neurodegeneration



Mean predicted trajectories of $A\beta$ [^{18}F]-florbetapir and [^{18}F]-fluorodeoxyglucose (FDG) positron emission tomography (PET), as well as magnetic resonance imaging (MRI)-derived hippocampal volume according to CSF AT status (only in Alzheimer's Disease Neuroimaging Initiative [ADNI]) are shown for individuals who were cognitively unimpaired (CU; A) or had mild cognitive impairment (MCI; B) at baseline. The mean predicted trajectories for

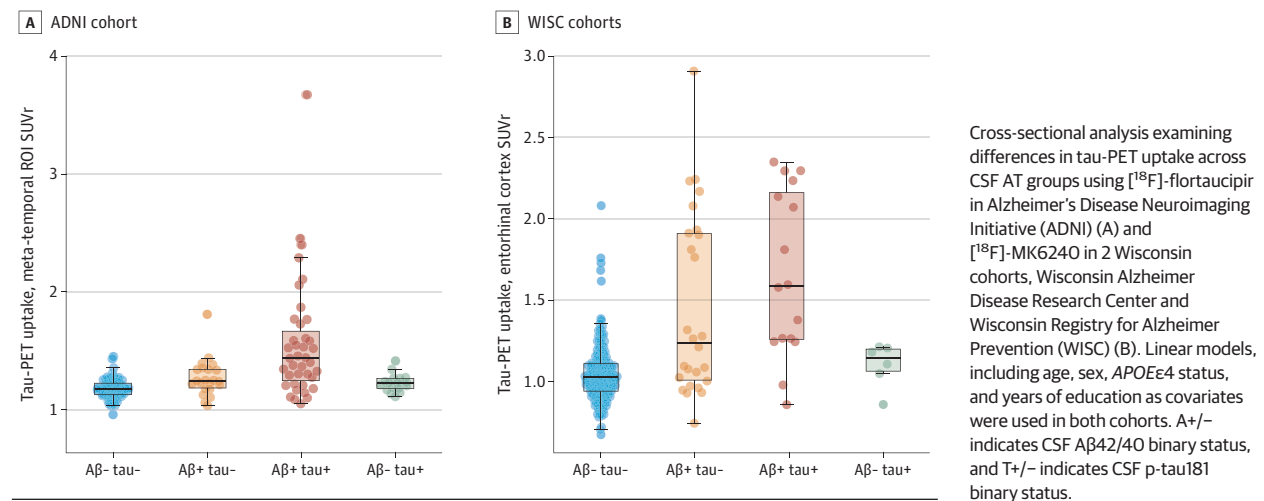
the $A\beta$ -PET and FDG-PET, as well as hippocampal volume are displayed with solid lines and 95% CIs. Trajectories were estimated including terms for CSF AT status, covariates (age, years of education, *APOE* $\epsilon 4$ genotype, sex), and the age \times CSF AT status interaction. A+/- indicates CSF $A\beta 42/40$ binary status, and T+/- indicates CSF p-tau181 binary status.

brain amyloidosis reflects slower CSF turnover. As an example of how alterations in CSF dynamics can impact biomarker readings, low concentrations of all core biomarkers (ie, p-tau, t-tau, $A\beta 42$, and $A\beta 40$) are commonly seen in cases of idiopathic normal pressure hydrocephalus, not necessarily associated with AD pathophysiology.³⁹ Interestingly, in the UGOT and ADNI cohorts, concentrations of $A\beta 40$ and $A\beta 42$ were clearly higher in the A-T+ group, compared with the A-T- group. This could potentially suggest that the CSF A-T+ profile is indeed associated altered CSF dynamics, given that within 2 $A\beta$ -negative groups as defined by the $A\beta 42/A\beta 40$ ratio, it would not be expected that $A\beta 42$ and $A\beta 40$ values alone would differ between groups.

Data from both longitudinal cohorts suggest that individuals with an A-T+ pattern had similar cognitive and biomarker trajectories as compared with participants with a CSF A-T- profile. This aligns with recent findings in individuals with subjective cognitive decline that showed that the A-T+ group was not more likely to progress to MCI or dementia compared with the biomarker negative group when defining "A" using $A\beta$ PET or $A\beta 42$ and "T" using p-tau.^{7,40} However, when defining A-T+ with both $A\beta$ - and tau-PET, individuals with this profile show slightly greater risk of cognitive decline than biomarker-negative individuals.⁴¹

Lastly, the choice of cutoff values may influence the interpretation of abnormal results. For $A\beta 42/40$, this is likely of lim-

Figure 4. Cross-Sectional Differences in Tau Positron Emission Tomography (PET) Deposition by Cerebrospinal Fluid (CSF) Amyloid-Tau (AT) Status



ited relevance because it usually presents a bimodal distribution²³ and concordance with amyloid PET usually reaches over 90%,⁴² which in turn makes cutoff determination and interpretation uncomplicated. However, p-tau typically shows no such distribution and is less clearly linked with tau-PET status.⁴³ Here, we show that both p-tau cutoff values that optimize sensitivity and specificity, as well as higher, less sensitive cutoff values presented very similar results in our study.

Strengths and Limitations

Strengths of this study include the large sample size of participants with core CSF biomarkers of A β and soluble tau pathology, allowing us to derive estimated prevalence of these pathologies in a real-world, unselected clinical routine sample, for whom memory clinic physicians order most CSF tests. Further, our consistent finding that CSF A-T+ individuals do not show significant cognitive decline suggests that the findings are generalizable across different populations. Limitations include limited clinical information from the UGOT cohort, as the data are derived from a database used in laboratory practice. This precludes us from defining strict inclusion/

exclusion criteria and describing the clinical features of the patients included. Another potential limitation is that the longitudinal cohorts consisted primarily of self-identified White individuals. Finally, AT status in the WISC sample was based on CSF tests not necessarily coinciding with the cognitive baseline visit.

Conclusions

In this cohort study, we found the estimated prevalence of a CSF A-T+ profile to be 4.1% in a large, real-world, clinical routine data set. Further, individuals with this profile did not present with significantly higher rates of cognitive decline or AD biomarker progression than those of individuals with an A-T- profile. We suggest that practitioners encountering this pattern in daily clinical practice should interpret such a finding similarly to CSF biomarker-negative results during the diagnostic workup of patients with cognitive complaints, and unusually high CSF A β 42 and A β 40 concentrations could be a recognizable feature of such a profile.

ARTICLE INFORMATION

Accepted for Publication: May 5, 2023.

Published Online: July 31, 2023.

doi:10.1001/jamaneurol.2023.2338

Open Access: This is an open access article distributed under the terms of the [CC-BY License](#). © 2023 Erickson P et al. *JAMA Neurology*.

Author Affiliations: Institute of Neuroscience and Physiology, Department of Psychiatry and Neurochemistry, Sahlgrenska Academy, University of Gothenburg, Gothenburg, Sweden (Erickson, Simrén, Brum, Ashton, Blennow, Andreasson, Zetterberg); Clinical Neurochemistry Laboratory, Sahlgrenska University Hospital, Gothenburg, Sweden (Simrén, Blennow, Andreasson, Zetterberg); Graduate Program in Biological Sciences: Biochemistry, Universidade Federal do Rio Grande do Sul, Porto Alegre, Brazil (Brum);

School of Medicine and Public Health, University of Wisconsin-Madison, Madison (Ennis, Langhough, Jonaitis, Van Hulle, Betthausen, Carlsson, Asthana, Johnson, Bendlin, Zetterberg); Wisconsin Alzheimer's Disease Research Center, University of Wisconsin-Madison School of Medicine and Public Health, Madison (Ennis, Langhough, Jonaitis, Van Hulle, Betthausen, Carlsson, Asthana, Johnson); Roche Diagnostics GmbH, Penzberg, Germany (Kollmorgen, Suridjan); Division of Geriatrics and Gerontology, Department of Medicine, University of Wisconsin-Madison School of Medicine and Public Health, Madison (Carlsson, Asthana); Geriatric Research Education and Clinical Center of the Wm. S. Middleton Memorial Veterans Hospital, Madison, Wisconsin (Carlsson, Asthana); Institute of Psychiatry, Psychology and Neuroscience, Maurice Wohl Institute Clinical Neuroscience Institute, King's College London, London, England

(Ashton); NIHR Biomedical Research Centre for Mental Health and Biomedical Research Unit for Dementia at South London and Maudsley NHS Foundation, London, England (Ashton); Centre for Age-Related Medicine, Stavanger University Hospital, Stavanger, Norway (Ashton); Department of Pathology and Laboratory Medicine, University of Pennsylvania School of Medicine, Philadelphia (Shaw); Institute of Neurology, Department of Neurodegenerative Disease, University College London, London, England (Zetterberg); UK Dementia Research Institute, University College London, London, England (Zetterberg); Hong Kong Center for Neurodegenerative Diseases, Hong Kong, China (Zetterberg).

Author Contributions: Drs Bendlin and Zetterberg had full access to all of the data in the study and take responsibility for the integrity of the data and the accuracy of the data analysis. Mr Erickson,

Dr Simrén, and Mr Brum contributed equally as co-first authors.

Concept and design: Erickson, Simrén, Brum, Ennis, Carlsson, Asthana, Andreasson, Bendlin, Zetterberg.

Acquisition, analysis, or interpretation of data: Erickson, Simrén, Brum, Ennis, Kollmorgen, Suridjan, Langhough, Jonaitis, Van Hulle, Betthausen, Carlsson, Ashton, Johnson, Shaw, Blennow, Bendlin, Zetterberg.

Drafting of the manuscript: Erickson, Simrén, Brum, Ennis, Ashton.

Critical revision of the manuscript for important intellectual content: Erickson, Simrén, Brum, Ennis, Kollmorgen, Suridjan, Langhough, Jonaitis, Van Hulle, Betthausen, Carlsson, Asthana, Johnson, Shaw, Blennow, Andreasson, Bendlin, Zetterberg.

Statistical analysis: Erickson, Simrén, Brum, Ennis, Jonaitis, Andreasson.

Obtained funding: Johnson, Blennow, Bendlin, Zetterberg.

Administrative, technical, or material support: Kollmorgen, Betthausen, Asthana, Shaw.

Supervision: Brum, Ashton, Blennow, Andreasson, Bendlin, Zetterberg.

Conflict of Interest Disclosures: Dr Ennis reported receiving grants from National Institute on Aging during the conduct of the study. Dr Kollmorgen reported being a full-time employee of Roche Diagnostics GmbH during the conduct of the study. Dr Langhough reported receiving grants from National Institutes of Health during the conduct of the study. Dr Jonaitis reported receiving grants from National Institutes of Health during the conduct of the study. Dr Betthausen reported receiving grants from the University of Wisconsin and National Institutes of Health and speaker honoraria from Intermountain Healthcare outside the submitted work. Dr Carlsson reported receiving grants from the National Institutes of Health, Eisai, Eli Lilly, Veterans Affairs; nonfinancial support from Amarin; data safety monitoring board/travel/ advisory board honoraria from Alzheimer's Association, National Institutes of Health, and American Fed Aging Res Beeson Program outside the submitted work. Dr Asthana reported receiving grants from National Institute on Aging/National Institutes of Health, Genentech, Merck, Toyoma Chemical, and Lundbeck outside the submitted work. Dr Johnson reported receiving grants from the National Institutes of Health and Cerveau and consultant fees from Roche Diagnostics, Merck, and ALZPath outside the submitted work. Dr Shaw reported receiving grants from the National Institute on Aging and teaching honoraria from Biogen outside the submitted work. Dr Blennow reported serving as a consultant and at advisory boards for Acumen, ALZPath, BioArctic, Biogen, Eisai, Julius Clinical, Lilly, Novartis, Ono Pharma, Prothena, Roche Diagnostics, and Siemens Healthineers; serving at data monitoring committees for Julius Clinical and Novartis; giving lectures, producing educational materials, and participating in educational programs for Biogen, Eisai, and Roche Diagnostics; and being a co-founder of Brain Biomarker Solutions in Gothenburg AB (BBS), which is a part of the GU Ventures Incubator Program, outside the work. Dr Bendlin reported receiving grants from the National Institutes of Health during the conduct of the study. Dr Zetterberg reported receiving personal/advisory board fees from AbbVie, Acumen, Alector, Alzinova, ALZPath, Annexon, Apellis, Artery Therapeutics,

AZTherapies, CogRx, Denali, Eisai, Nervgen, Novo Nordisk, Optoceutics, Passage Bio, Pinteon Therapeutics, Prothena, Red Abbey Labs, reMYND, Roche, Samumed, Siemens Healthineers, Triplet Therapeutics, and Wave Scientific; receiving lecture fees from Collectricron, Fujirebio, Alzecure, Biogen, and Roche outside the submitted work; and being a co-founder of Brain Biomarker Solutions in Gothenburg AB (BBS), which is a part of the GU Ventures Incubator Program. No other disclosures were reported.

Funding/Support: This study was supported in part by Gunn och Bertil Stohnes Stiftelse, the Swedish Dementia Foundation (Demensfonden), Sahlgrenska University funds, Göteborgs Läkaresällskap (Gothenburg Physicians' Association), and Stiftelsen för Gamla Tjänarinnor (Dr Simrén); the Wallenberg Centre for Molecular and Translational Medicine, Swedish Alzheimer Foundation, Hjärnfonden Sweden, the Swedish Dementia Foundation (Demensfonden), and Stiftelsen för Gamla Tjänarinnor (Dr Ashton); grant 2017-00915 from the Swedish Research Council, grant RDAPB-201809-2016615 from the Alzheimer Drug Discovery Foundation, grant AF-742881 from the Swedish Alzheimer Foundation, grant FO2017-0243 from Hjärnfonden Sweden, the Swedish state under the agreement between the Swedish government and the County Councils, grant ALFGBG-715986 from the Avtal om Läkarutbildning och Forskning agreement, grant JPND2019-466-236 from the European Union Joint Program for Neurodegenerative Disorders, and grant IRO1AGO68398-01 from the National Institutes of Health (Dr Blennow); grant 2022-01018 from the Swedish Research Council, the European Union's Horizon Europe research and innovation programme under grant agreement No. 101053962, grant ALFGBG-71320 from the Swedish State Support for Clinical Research, the Alzheimer Drug Discovery Foundation, grant 201809-2016862 from USA, grants ADSF-21-831376-C, ADSF-21-831381-C, and ADSF-21-831377-C from the AD Strategic Fund and the Alzheimer's Association, the Bluefield Project, the Olav Thon Foundation, the Erling-Persson Family Foundation, grant FO2022-0270 from Stiftelsen för Gamla Tjänarinnor, Hjärnfonden Sweden, the European Union's Horizon 2020 research and innovation programme under the Marie Skłodowska-Curie grant agreement No. 860197 (MIRIAD), grant JPND2021-00694 from the European Union Joint Programme—Neurodegenerative Disease Research, and grant UKDRI-1003 from the UK Dementia Research Institute at UCL (Dr Zetterberg, Wallenberg Scholar); grants RO1AGO70883, RO1AGO62285, P30AGO62715, RO1AGO59312, RO1AGO70973, and RO1AGO37639 from the National Institute on Aging (Dr Bendlin); grant U19 AG024904-QC from the National Institute on Aging, grant AG072979 from Roche, and grant 13637.01 from Michael J. Fox Foundation for Parkinson's Research (Dr Shaw); and the WISC cohorts were funded with grants RO1AGO27161, RO1AGO21155, and P30AGO62715 from the National Institutes of Health.

Role of the Funder/Sponsor: The funders had no role in the design and conduct of the study; collection, management, analysis, and interpretation of the data; preparation, review, or approval of the manuscript; and decision to submit the manuscript for publication.

Group Information: The members of the ADNI Cohort appear in [Supplement 2](#).

Data Sharing Statement: See [Supplement 3](#).

REFERENCES

1. Dubois B, Villain N, Frisoni GB, et al. Clinical diagnosis of Alzheimer disease: recommendations of the International Working Group. *Lancet Neurol*. 2021;20(6):484-496. doi:10.1016/S1474-4422(21)00066-1
2. Jack CR Jr, Bennett DA, Blennow K, et al; Contributors. NIA-AA Research Framework: toward a biological definition of Alzheimer disease. *Alzheimers Dement*. 2018;14(4):535-562. doi:10.1016/j.jalz.2018.02.018
3. Zetterberg H, Bendlin BB. Biomarkers for Alzheimer disease—preparing for a new era of disease-modifying therapies. *Mol Psychiatry*. 2021;26(1):296-308. doi:10.1038/s41380-020-0721-9
4. Itoh N, Arai H, Urakami K, et al. Large-scale, multicenter study of cerebrospinal fluid tau protein phosphorylated at serine 199 for the antemortem diagnosis of Alzheimer disease. *Ann Neurol*. 2001;50(2):150-156. doi:10.1002/ana.1054
5. Skillbäck T, Farahmand BY, Rosén C, et al. Cerebrospinal fluid tau and amyloid- β 1-42 in patients with dementia. *Brain*. 2015;138(Pt 9):2716-2731. doi:10.1093/brain/awv181
6. Jack CR Jr, Knopman DS, Chételat G, et al. Suspected non-Alzheimer disease pathophysiology—concept and controversy. *Nat Rev Neurol*. 2016;12(2):117-124. doi:10.1038/nrneurol.2015.251
7. Eibenau JL, Timmers T, Wesselman LMP, et al. ATN classification and clinical progression in subjective cognitive decline: the SCIENCE project. *Neurology*. 2020;95(1):e46-e58. doi:10.1212/WNL.00000000000009724
8. Kern S, Zetterberg H, Kern J, et al. Prevalence of preclinical Alzheimer disease: comparison of current classification systems. *Neurology*. 2018;90(19):e1682-e1691. doi:10.1212/WNL.00000000000005476
9. Soldan A, Pettigrew C, Fagan AM, et al. ATN profiles among cognitively normal individuals and longitudinal cognitive outcomes. *Neurology*. 2019;92(14):e1567-e1579. doi:10.1212/WNL.00000000000007248
10. Vos SJ, Xiong C, Visser PJ, et al. Preclinical Alzheimer disease and its outcome: a longitudinal cohort study. *Lancet Neurol*. 2013;12(10):957-965. doi:10.1016/S1474-4422(13)70194-7
11. Blennow K, Shaw LM, Stomrud E, et al. Predicting clinical decline and conversion to Alzheimer disease or dementia using novel Elecsys A β (1-42), pTau and tTau CSF immunoassays. *Sci Rep*. 2019;9(1):19024. doi:10.1038/s41598-019-54204-z
12. Guo T, Korman D, La Joie R, et al; Alzheimer's Disease Neuroimaging Initiative. Normalization of CSF pTau measurement by A β ₄₀ improves its performance as a biomarker of Alzheimer disease. *Alzheimers Res Ther*. 2020;12(1):97. doi:10.1186/s13195-020-00665-8
13. Fleisher AS, Pontecorvo MJ, Devous MD Sr, et al; A16 Study Investigators. Positron emission tomography imaging with [18F]flortaucipir and postmortem assessment of Alzheimer disease neuropathologic changes. *JAMA Neurol*. 2020;77(7):829-839. doi:10.1001/jamaneurol.2020.0528

14. Pascoal TA, Benedet AL, Tudorascu DL, et al. Longitudinal 18F-MK-6240 tau tangles accumulation follows Braak stages. *Brain*. 2021;144(11):3517-3528. doi:10.1093/brain/awab248
15. Kaeser SA, Häslér LM, Lambert M, et al. CSF p-tau increase in response to A β -type and Danish-type cerebral amyloidosis and in the absence of neurofibrillary tangles. *Acta Neuropathol*. 2022;143(2):287-290. doi:10.1007/s00401-021-02400-5
16. Pichet Binette A, Franzmeier N, Spotorno N, et al; Alzheimer's Disease Neuroimaging Initiative. Amyloid-associated increases in soluble tau relate to tau aggregation rates and cognitive decline in early Alzheimer disease. *Nat Commun*. 2022;13(1):6635. doi:10.1038/s41467-022-34129-4
17. Mattsson-Carlgen N, Andersson E, Janelidze S, et al. A β deposition is associated with increases in soluble and phosphorylated tau that precede a positive tau PET in Alzheimer disease. *Sci Adv*. 2020;6(16):eaz2387. doi:10.1126/sciadv.aaz2387
18. Sato C, Barthélemy NR, Mawuenyega KG, et al. Tau kinetics in neurons and the human central nervous system. *Neuron*. 2018;97(6):1284-1298.e7. doi:10.1016/j.neuron.2018.02.015
19. Yasuhara O, Kawamata T, Aimi Y, McGeer EG, McGeer PL. Two types of dystrophic neurites in senile plaques of Alzheimer disease and elderly non-demented cases. *Neurosci Lett*. 1994;171(1-2):73-76. doi:10.1016/0304-3940(94)90608-4
20. Su JH, Cummings BJ, Cotman CW. Early phosphorylation of tau in Alzheimer disease occurs at ser-202 and is preferentially located within neurites. *Neuroreport*. 1994;5(17):2358-2362. doi:10.1097/00001756-199411000-00037
21. Johnson SC, Kosik RL, Jonaitis EM, et al. The Wisconsin Registry for Alzheimer Prevention: a review of findings and current directions. *Alzheimers Dement (Amst)*. 2017;10:130-142. doi:10.1016/j.dadm.2017.11.007
22. Alzheimer's Disease Neuroimaging Initiative. Home page. Accessed April 17, 2023. <https://adni.loni.usc.edu/>
23. Gobom J, Parnetti L, Rosa-Neto P, et al. Validation of the LUMIPULSE automated immunoassay for the measurement of core AD biomarkers in cerebrospinal fluid. *Clin Chem Lab Med*. 2021;60(2):207-219. doi:10.1515/ccml-2021-0651
24. Jagust WJ, Landau SM, Koeppe RA, et al. The Alzheimer's Disease Neuroimaging Initiative 2 PET core: 2015. *Alzheimers Dement*. 2015;11(7):757-771. doi:10.1016/j.jalz.2015.05.001
25. Wyman BT, Harvey DJ, Crawford K, et al; Alzheimer's Disease Neuroimaging Initiative. Standardization of analysis sets for reporting results from ADNI MRI data. *Alzheimers Dement*. 2013;9(3):332-337. doi:10.1016/j.jalz.2012.06.004
26. Betthausen TJ, Cody KA, Zammit MD, et al. In vivo characterization and quantification of neurofibrillary tau PET radioligand ¹⁸F-MK-6240 in humans from Alzheimer disease dementia to young controls. *J Nucl Med*. 2019;60(1):93-99. doi:10.2967/jnumed.118.209650
27. Jack CR Jr, Wiste HJ, Weigand SD, et al. Defining imaging biomarker cut points for brain aging and Alzheimer disease. *Alzheimers Dement*. 2017;13(3):205-216. doi:10.1016/j.jalz.2016.08.005
28. Donohue MC, Sperling RA, Salmon DP, et al; Australian Imaging, Biomarkers, and Lifestyle Flagship Study of Ageing; Alzheimer's Disease Neuroimaging Initiative; Alzheimer's Disease Cooperative Study. The preclinical Alzheimer cognitive composite: measuring amyloid-related decline. *JAMA Neurol*. 2014;71(8):961-970. doi:10.1001/jamaneurol.2014.803
29. Jonaitis EM, Kosik RL, Clark LR, et al. Measuring longitudinal cognition: individual tests versus composites. *Alzheimers Dement (Amst)*. 2019;11:74-84. doi:10.1016/j.dadm.2018.11.006
30. Monsell SE, Dodge HH, Zhou XH, et al; Neuropsychology Work Group Advisory to the Clinical Task Force. Results from the NACC uniform data set Neuropsychological Battery Crosswalk Study. *Alzheimer Dis Assoc Disord*. 2016;30(2):134-139. doi:10.1097/WAD.0000000000000111
31. Mattsson-Carlgen N, Janelidze S, Palmqvist S, et al. Longitudinal plasma p-tau217 is increased in early stages of Alzheimer disease. *Brain*. 2020;143(11):3234-3241. doi:10.1093/brain/awaa286
32. Brum WS, de Bastiani MA, Bieger A, et al; Alzheimer's Disease Neuroimaging Initiative (ADNI). A three-range approach enhances the prognostic utility of CSF biomarkers in Alzheimer disease. *Alzheimers Dement (N Y)*. 2022;8(1):e12270. doi:10.1002/trc2.12270
33. Jansen WJ, Janssen O, Tijms BM, et al; Amyloid Biomarker Study Group. Prevalence estimates of amyloid abnormality across the Alzheimer disease clinical spectrum. *JAMA Neurol*. 2022;79(3):228-243. doi:10.1001/jamaneurol.2021.5216
34. Rosenberg A, Öhlund-Wistbacka U, Hall A, et al. β -Amyloid, tau, neurodegeneration classification and eligibility for anti-amyloid treatment in a memory clinic population. *Neurology*. 2022;99(19):e2102-e2113. doi:10.1212/WNL.00000000000021043
35. Mattsson-Carlgen N, Leuzy A, Janelidze S, et al. The implications of different approaches to define AT(N) in Alzheimer disease. *Neurology*. 2020;94(21):e2233-e2244. doi:10.1212/WNL.0000000000009485
36. Hansson O, Lehmann S, Otto M, Zetterberg H, Lewczuk P. Advantages and disadvantages of the use of the CSF amyloid β (A β) 42/40 ratio in the diagnosis of Alzheimer disease. *Alzheimers Res Ther*. 2019;11(1):34. doi:10.1186/s13195-019-0485-0
37. Palmqvist S, Janelidze S, Quiroz YT, et al. Discriminative accuracy of plasma phospho-tau217 for Alzheimer disease vs other neurodegenerative disorders. *JAMA*. 2020;324(8):772-781. doi:10.1001/jama.2020.12134
38. Mattsson-Carlgen N, Janelidze S, Bateman RJ, et al. Soluble p-tau217 reflects amyloid and tau pathology and mediates the association of amyloid with tau. *EMBO Mol Med*. 2021;13(6):e14022. doi:10.15252/emmm.202114022
39. Jeppsson A, Wikkelsö C, Blennow K, et al. CSF biomarkers distinguish idiopathic normal pressure hydrocephalus from its mimics. *J Neurol Neurosurg Psychiatry*. 2019;90(10):1117-1123. doi:10.1136/jnnp-2019-320826
40. Oberstein TJ, Schmidt MA, Florvaag A, et al. Amyloid- β levels and cognitive trajectories in nondemented pTau181-positive subjects without amyloidopathy. *Brain*. 2022;145(11):4032-4041. doi:10.1093/brain/awac297
41. Jack CR Jr, Wiste HJ, Thorneau TM, et al. Associations of amyloid, tau, and neurodegeneration biomarker profiles with rates of memory decline among individuals without dementia. *JAMA*. 2019;321(23):2316-2325. doi:10.1001/jama.2019.7437
42. Willemse EAJ, Tijms BM, van Berckel BNM, et al. Comparing CSF amyloid- β biomarker ratios for 2 automated immunoassays, elecsys and lumipulse, with amyloid PET status. *Alzheimers Dement (Amst)*. 2021;13(1):e12182. doi:10.1002/dad2.12182
43. Ossenkoppele R, Reimand J, Smith R, et al; Alzheimer's Disease Neuroimaging Initiative. Tau PET correlates with different Alzheimer disease-related features compared to CSF and plasma p-tau biomarkers. *EMBO Mol Med*. 2021;13(8):e14398. doi:10.15252/emmm.202114398